

## New pentacyclic polyketide dimeric peroxides from a Taiwanese marine sponge *Petrosia elastica*

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**Abstract**—Two novel pentacyclic polyketide dimers, dihalenaquinolides A (**1**) and B (**2**), have been isolated from the marine sponge *Petrosia elastica* collected in Nan-wan, Taiwan. The structures **1** and **2** were established on the basis of extensive spectral analysis.

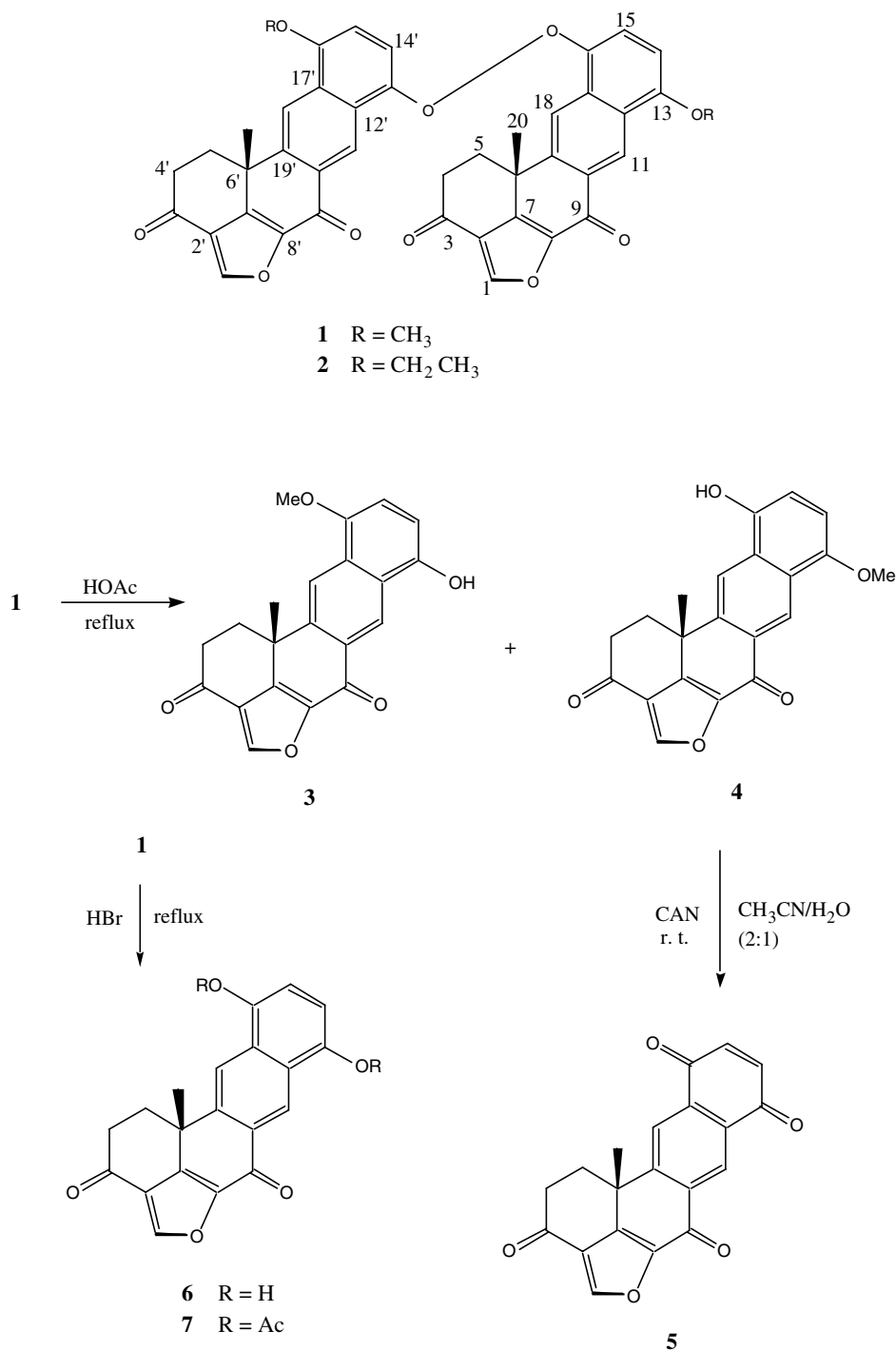
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Tropical marine sponges are a fertile source of secondary metabolites with diverse and often novel molecular architecture.<sup>1</sup> In the course of our study on biologically active secondary metabolites from marine organisms,<sup>2</sup> we have undertaken the chemical examination of the Taiwanese marine sponge *Petrosia elastica* Keller.<sup>3</sup> The sponge material (1.85 kg) was extracted exhaustively with EtOH (31 × 3) to give a residue (27.3 g), which was partitioned between water (500 mL) and EtOAc (11 × 3) furnishing the EtOAc residue (12.6 g). Chromatography of the residue over silica gel with solvents of increasing polarity from *n*-hexane/CHCl<sub>3</sub> through CHCl<sub>3</sub> to CHCl<sub>3</sub>/MeOH gave six fractions (A–F). Fraction F on further chromatography over silica gel followed by preparative TLC (CHCl<sub>3</sub>/MeOH, 95:5) furnished dihalenaquinolides A (**1**) and B (**2**).

Dihalenaquinolide A (**1**) has a formula C<sub>42</sub>H<sub>30</sub>O<sub>10</sub> as deduced from LRFAB and HRFAB-MS spectra. The FAB-MS showed a quasi-molecular ion peak at *m/z* 695 ([M+H]<sup>+</sup>) together with a strong mass fragment at *m/z* 348 corresponding to the fragmentation of its constituent monomer (C<sub>21</sub>H<sub>16</sub>O<sub>5</sub>), suggesting that compound **1** is a dimer. Its <sup>1</sup>H NMR spectrum showed the presence of four doublets at δ 6.65, 6.75, 6.85, and 6.95 each having a coupling constant 8 Hz, four singlets at δ 8.21, 8.26, 9.22 and 9.26, and a singlet for two methyls at δ 1.63, which is similar to the <sup>1</sup>H NMR spectrum of

halenaquinol except for the presence of two sets of protons in compound **1** against one in halenaquinol.<sup>4</sup> Methylation of **1** with CH<sub>2</sub>N<sub>2</sub> was not successful. This indicated the absence of aromatic hydroxyl group in its structure. Further its <sup>1</sup>H NMR spectrum showed the presence of a singlet for two methyls at δ 3.95 suggesting that the hydroxyl group in halenaquinol has been replaced by methoxyl group in compound **1**, which was further supported by the presence of methoxyl carbon at δ 55.7 in <sup>13</sup>C NMR spectrum. The presence of peroxide linkage was supported by the strong mass fragments at *m/z* 348 ([M+H–C<sub>21</sub>H<sub>15</sub>O<sub>5</sub>]<sup>+</sup>) and 333 ([M–C<sub>21</sub>H<sub>14</sub>O<sub>6</sub>]<sup>+</sup>) in the mass spectrum. The nature of the rings A, B, C, and D in the two monomers was observed symmetrical, which led to the doubling of signals or with a very small difference in its <sup>13</sup>C NMR values.<sup>5</sup> The similar nature of the two monomers was also supported by the presence of a singlet for two protons H-18/H-18' at δ 8.21, another singlet for two protons H-1/H-1' at δ 8.26, and the two singlets at δ 9.22 and 9.26 having a very small difference in their δ value can be assignable to H-11 or H-11' in its <sup>1</sup>H NMR spectrum. The distortion to the symmetry was observed in ring E from C-13/C-13' to C-16/C-16'. This inferred the only possibility for the unsymmetrical attachment of the two monomers at C-16 and C-13' as shown in compound **1** and the positions of the methoxyl groups at C-13 and C-16'. NOESY spectrum showed the correlation between the methoxyl and the protons H-11 at δ 9.22 and H-18' at δ 8.21, respectively, confirming further the unsymmetrical nature of the two monomers. A close comparison of the <sup>13</sup>C NMR values of halenaquinol and **1** revealed that the two carbons, C-13 and C-16' in **1** are downfield by 1.3 and 1.7 ppm, respectively, which can be explainable by the

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presence of methoxyl groups at those positions. The <sup>1</sup>H NMR data at δ 6.65 and 6.95 were assigned to H-14 and H-15, respectively, on the basis of COSY (H-14/H-15), HMBC (H-14/C-13, C-15, C-16; H-15/C-14, C-16), and NOESY spectral data. Similarly the other two protons at δ 6.85 and 6.75 were assigned to H-14' and H-15', respectively. Furthermore, the assignment of C-13, C-13', C-16 and C-16' at δ 148.6, 147.1, 145.5, and 147.2 was completed by HMBC too. Dihalenaquinolide A (**1**, 4 mg) was refluxed in acetic acid (4 mL) to give **3** and **4**, which were further reacted with CAN (ceric ammonium nitrate) in CH<sub>3</sub>CN/H<sub>2</sub>O (2:1) under room temperature

to yield helenoquinone (**5**, 1.4 mg).<sup>6</sup> When compound **1** was refluxed with HBr (48%, 1 mL) in AcOH for 12 h, helenoquinol (**6**) was produced and was isolated as its diacetate form (**7**). The spectral data (<sup>1</sup>H NMR, EI, and FAB-MS) of the known **5–7** are identical with those of authentic samples, respectively.<sup>7</sup> The new derivatives **3** and **4**, which exhibited more polar than **1**, were obtained as an inseparable diastereomeric mixture.<sup>8</sup>

Dihalenaquinolide B (**2**) has been deduced as C<sub>44</sub>H<sub>34</sub>O<sub>10</sub> from HRFAB-MS and DEPT spectrum. Its <sup>1</sup>H NMR spectrum showed the presence of four doublets at δ 6.64,

**Table 1.** Cytotoxicity of polycyclic hydroquinones and quinones against human tumor cells (growth inhibition, %)<sup>a</sup>

Compound (10 µg/mL)	PC-3 <sup>b</sup>	Hep3B <sup>c</sup>
Dihalenaquinolide A ( <b>1</b> )	56	5
Dihalenaquinolide B ( <b>2</b> )	35	5
Halenaquinone ( <b>5</b> )	60	47
Halenaquinol ( <b>6</b> )	98	20
Halenaquinol diacetate ( <b>7</b> )	43	31
Xestoquinone	100	2
Xestoquinolide	50	5
Taxol (0.1 µg/mL)	80	85

<sup>a</sup> Positive compound: % of control >50%.<sup>b</sup> PC-3: human prostate cancer cells.<sup>c</sup> Hep3B: human hepatoma cancer cells.

6.77, 6.86, 6.94 corresponds to H-14, H-15, H-15', H-14', respectively, three singlets at  $\delta$  8.21 (H-18/H-18'), 8.26 (H-1/H-1') and 9.24 (H-11/H-11'), and a singlet for two methyls at  $\delta$  1.64 similar to that of **1** except the presence of a quartet for two methylene groups at  $\delta$  4.14 ( $J = 6.9$  Hz) and a triplet for two methyl groups at  $\delta$  1.52 ( $J = 6.6$  Hz) in place of the singlet for methoxyl group at  $\delta$  3.95 in **1**. From its EIMS it was observed that **2** is 28 mass units greater than **1** suggests that compound **2** might be having ethoxyl group at C-13 instead of methoxyl group of **1**. This was further supported the presence of the two <sup>13</sup>C NMR signals at  $\delta$  64.2 and 14.9 for OCH<sub>2</sub>CH<sub>3</sub>. The <sup>13</sup>C NMR values were assigned for each carbon and were in good agreement with the structure.<sup>9</sup>

The cytotoxic activity of the isolated pentacyclic hydroquinones and quinones were tested in vitro against human prostate (PC-3) and hepatoma (Hep3B) tumor cell lines. As shown in Table 1, compounds **1**, **5**, **6**, and xestoquinone selectively inhibited the growth of PC-3 tumor cells at 10 µg/mL, while compounds **2**, **7**, and xestoquinolide were inactive. None of the compounds showed growth inhibition toward human Hep3B tumor cells.

**Cytotoxicity Assay.** The bioassay used against PC-3 (human androgen-independent prostate carcinoma) and Hep3B (human hepatocellular carcinoma) tumor cells was based on a sulforhodamine B (SRB) assay method.<sup>10</sup> The procedure of assay was carried out as previously described.<sup>11</sup> Taxol was used as a standard compound.

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- Dihalenaquinolide A:  $[\alpha]_D^{25} +45^\circ$  (*c* 0.1, CHCl<sub>3</sub>); UV (MeOH)  $\lambda_{max}$  229.4, 300.6 nm; IR (CHCl<sub>3</sub>)  $\nu_{max}$  3104, 3035, 1727, 1685, 1523, 1427, 1018, 995, 767, and 667 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.26 (s, 2H, H-1/H-1'), 2.8–3.0 (m, 4H, H-4/H-4'), 2.28 and 2.76 (m, 4H, H-5/H-5'), 9.22 (s, <sup>1</sup>H, H-11), 9.26 (<sup>1</sup>H, H-11'), 6.65 (d,  $J = 8$  Hz, <sup>1</sup>H, H-14), 6.85 (d,  $J = 8$  Hz, <sup>1</sup>H, H-14'), 6.95 (d,  $J = 8$  Hz, <sup>1</sup>H, H-15), 6.75 (d,  $J = 8$  Hz, <sup>1</sup>H, H-15'), 8.21 (2H, H-18/H-18'), 1.63 (s, 6H, H-20/H-20'), and 3.95 (s, 6H, 2 × OCH<sub>3</sub>); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  150.9 (C-1/C-1'), 122.4 (C-2/C-2'), 192.3, 192.4 (C-3, C-3'), 36.7 (C-4/C-4'), 34.0 (C-5/C-5'), 35.7 (C-6/C-6'), 147.9 (C-7/C-7'), 144.7 (C-8/C-8'), 172.8, 172.9 (C-9, C-9'), 130.0 (C-10/C-10'), 123.8 (C-11/C-11'), 122.4 (C-12), 123.8 (C-12'), 148.6 (C-13), 147.1 (C-13'), 104.1 (C-14), 109.0 (C-14'), 111.4 (C-15), 107.1 (C-15'), 145.5 (C-16), 147.2 (C-16'), 124.6 (C-17), 124.8 (C-17'), 118.5, 118.7 (C-18, C-18'), 147.5 (C-19/C-19'), 31.9 (C-20/C-20'), and 55.7 (OCH<sub>3</sub>); FAB-MS *m/z*: 695 ([M+H]<sup>+</sup>, 0.7), 694 (M<sup>+</sup>, 0.5), 349 ([M-C<sub>21</sub>H<sub>13</sub>O<sub>5</sub>]<sup>+</sup>, 23.6), 348 ([M-C<sub>21</sub>H<sub>14</sub>O<sub>5</sub>]<sup>+</sup>, 17.4), 334 ([M-C<sub>21</sub>H<sub>13</sub>O<sub>6</sub>]<sup>+</sup>, 6.2), 333 ([M-C<sub>21</sub>H<sub>14</sub>O<sub>6</sub>]<sup>+</sup>, 6.1), 307 (4.93), 289 (4.85), 154 (88.6), and 136 (88.5); HRFAB-MS *m/z*: [M+H]<sup>+</sup> 695.1912 (calcd for C<sub>42</sub>H<sub>31</sub>O<sub>10</sub>, 695.1917); TLC *R<sub>f</sub>*: 0.70/CH<sub>2</sub>Cl<sub>2</sub>-MeOH (49:1), 0.53/CH<sub>2</sub>Cl<sub>2</sub>-MeOH (99:1).
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- Compounds **3** and **4**: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  9.24, 9.22 (s, 2H, H-1), 8.30, 8.27 (s, 2H, H-18), 8.23 (s, 2H, H-11), 6.66, 6.87 (d,  $J = 8.1$  Hz, 2H, H-14), 6.79, 6.91 (d,  $J = 8.1$  Hz, 2H, H-15), 3.98 (s, 6H, OCH<sub>3</sub>), 2.85 (m, 6H), 2.32 (m, 2H), 1.67 (s, 6H, H-20); EIMS (70 eV) *m/z* 348 (M<sup>+</sup>, 77), 333 ([M-Me]<sup>+</sup>, 100); FAB-MS *m/z*: 349 ([M+H]<sup>+</sup>, 3.2); TLC *R<sub>f</sub>*: 0.63/CH<sub>2</sub>Cl<sub>2</sub>-MeOH (49:1).
- Dihalenaquinolide B:  $[\alpha]_D^{25} +70.8^\circ$  (*c* 0.1, CHCl<sub>3</sub>); UV (MeOH)  $\lambda_{max}$  227.8, 299.8 nm; IR (CHCl<sub>3</sub>)  $\nu_{max}$  3135, 3058, 2904, 1677, 1608, 1106, 1025, 917, 767, and 671 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  8.21 (2H, H-1/H-1'), 2.80–2.98 (m, 4H, H-4/H-4'), 2.29 and 2.90 (m, 4H, H-5/H-5'), 9.24 (2H, H-11/H-11'), 6.64 (d,  $J = 8$  Hz, 1H, H-14), 6.86 (d,  $J = 8$  Hz, 1H, H-14'), 6.94 (d,  $J = 8$  Hz, 1H, H-15), 6.77 (d,  $J = 8$  Hz, 1H, H-15'), 8.26 (2H, H-18/H-18'), 1.64 (6H, H-20/H-20'), 4.14 (q,  $J = 6.9$  Hz, 4H, OCH<sub>2</sub>CH<sub>3</sub>), and 1.52 (t,  $J = 6.6$  Hz, 6H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  150.1 (C-1/C-1'), 122.4 (C-2/C-2'), 192.4, 192.5 (C-3/C-3'), 36.7 (C-4/C-4'), 34.0 (C-5/C-5'), 35.7 (C-6/C-6'), 144.0 (C-7/C-7'), 144.7 (C-8/C-8'), 172.8, 172.9 (C-9/C-9'), 129.9 (C-10/C-10'), 123.8 (C-11/C-11'), 122.4 (C-12/C-12'), 148.6 (C-13), 147.1 (C-13'), 105.1 (C-14), 109.1 (C-14'), 111.5 (C-15), 108.3 (C-15'), 145.5 (C-16), 147.2 (C-16'), 124.6 (C-17/C-17'), 118.4, 118.6 (C-18, C-18'), 147.5 (C-19/C-19'), 31.7 (C-20/C-20'), 64.1 (OCH<sub>2</sub>CH<sub>3</sub>), and 14.8 (OCH<sub>2</sub>CH<sub>3</sub>); FAB-MS *m/z*: 723

([M+H]<sup>+</sup>, 0.7), 722 (M<sup>+</sup>, 0.4), 363 ([M-C<sub>22</sub>H<sub>16</sub>O<sub>5</sub>]<sup>+</sup>, 49), 362 ([M-C<sub>22</sub>H<sub>17</sub>O<sub>5</sub>]<sup>+</sup>, 37), 348 (M-C<sub>22</sub>H<sub>15</sub>O<sub>6</sub>+H)<sup>+</sup>, 7.6), 347 ([M-C<sub>22</sub>H<sub>16</sub>O<sub>6</sub>]<sup>+</sup>, 6), 307 (8.3), 289 (6.7), 154 (90.6), and 136 (100); HRFAB-MS *m/z*: [M]<sup>+</sup> 722.2165 (calcd for C<sub>44</sub>H<sub>34</sub>O<sub>10</sub>, 722.2152); [M+H]<sup>+</sup> 723.2231 (calcd for C<sub>44</sub>H<sub>35</sub>O<sub>10</sub>, 723.2230); TLC *R<sub>f</sub>*: 0.58/CH<sub>2</sub>Cl<sub>2</sub>-MeOH (99:1).

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